101Bio

**Product Name:** Lentivirus 10X Titer-Up Reagent

**Cat. #:** P906 (1 ml) Cat.# at FisherSci.com is NC1792390 (1 ml), P906S (0.1 ml test sample)

Application: Increase lentivirus titer by up to **10 times** in virus packaging procedure.

> 6-14 hours after transfection of human embryonic kidney (HEK) 293T cells with retroviral or lentiviral packaging plasmid mix, replace the culture medium with fresh DMEM medium supplemented with 10% heat-

inactivated fetal bovine serum and 0.5% penicillin-streptomycin, and add 1/500 volume of ViralBoost Reagent to one volume of fresh culture medium and continue incubation in the CO2 incubator at 37°C. This

product is for research use only.

**Product Size:** P906: 1 ml (for packaging 500 mL lentivirus soup); P906S: 0.1 ml

**Product Description:** This product is a novel recipe of small molecules designed for effective

lentivirus packaging.

✓ Increase virus titer by up to 10 times

✓ Increase viral RNA transcription

✓ Increase virus particle packaging efficiency

Shipping / Storage: Ship at room temperature and store at 4°C

Shelf Life: 12 months

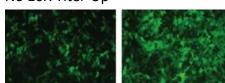
Remark: Each lot of 10X Titer-Up reagent is **functionally tested** in virus production

experiment using 293T cells. Follow the recommended NIH BSL-2

guidelines for all materials containing Lentivirus.

No 10X Titer-Up Use 10X Titer-Up

Fig. 1



HEK293 cells transduced by GFP lentivirus which was packaged with or without 10X Titer-Up Reagent.

# **Protocol** (example of producing virus in 100mm petri dish)

# Day 1: Plating cells

- 1. 24 hours before transfection, coat 100mm dishes with 6 mL 1x Gelatin for 60 min.
- 2. Aspirate gelatin, and plate ~5 X 10<sup>6</sup> fast growing 293T cells per plate, in 10 ml medium.

**Note**: Before plating, pipet the 293T cell suspension vigorously to achieve single cells. Plate the cells in evenly distribution manor.

### Day 2: Transfection (using 101Bio "293T Trasfection Reagent", Cat. #: P902.)

- 1. Change medium: 2 hours before transfection, remove culture medium and add 10 mL fresh complete culture medium (with 10% serum / without antibiotics)
- 2. In tube 1 add: ~20 μg DNA (virus vector and packaging mix [101Bio, Cat. #: P904P]) 500 μL DMEM (serum-free, High Glucose)

Pipet up and down to mix well

3. In tube 2 Add: 40 µL 293T Transfection Reagent for Lentivirus Packaging 500 µL DMEM (serum-free, High Glucose)

Gently pipet up and down to mix well

- 4. Incubate at room temperature (20 ~ 25°C) for 3 min.
- 5. Add tube 2 into tube 1, pipet up and down several times. **Vortex** for **10 seconds**.
- 6. Incubate for **15 minutes** at **room temperature**.
- 7. Add the incubated mixture drop-wise to the cells, and gently rock / swirl the plate.
- 8. Return the cells to 37°C incubator with 5% CO<sub>2</sub>.

# Day 3: Add 10X Titer-Up

Add 20 µl of 10X Titer-Up (500x) to the medium. Return the plates to the cell culture incubator.

#### Day 4: Collect virus

- 1. Collect virus supernatant twice at **48 and 72 hours** post transfection into a 50mL sterile FALCON tube. Centrifuge at **3,000 rpm** for **15 minutes** at 4°C to remove cell debris. Filter the clear supernatant through **0.45 μm** syringe tip filter.
- 2. The filtered clear supernatant is virus soup. Use it immediately or aliquot into sterile 1.5-mL tubes and store at **-80°C**, for **up to 3 months**.
- 3. (Optional) To concentrate virus, add 1 volume of **5X Lentivirus Concentration Solution** (101Bio, Cat. #: P904C, Cat.# at FisherSci.com is NC3242189) to 4 volume of the viral supernatant (volume of Lentivirus Concentration Solution vs. volume of viral supernatant = 1:4) and mix thoroughly. Put the mixture to the 4°C refrigerator overnight and spin the virus pellet down next day. Please refer to our P904C user manual for details.
- 4. **Note:** No side effect 10X Titer-Up reagent on gene expression has been detected when directly transduce 293T cells. It may be various on different cell types (cell lines). A pilot test on the side effect of 10X Titer-Up is advised on sensitive cells.

California, USA